

# Engineering Promiscuity of Chloramphenicol Acetyltransferase for Microbial Designer Ester Biosynthesis



U.S. DEPARTMENT OF  
**ENERGY**

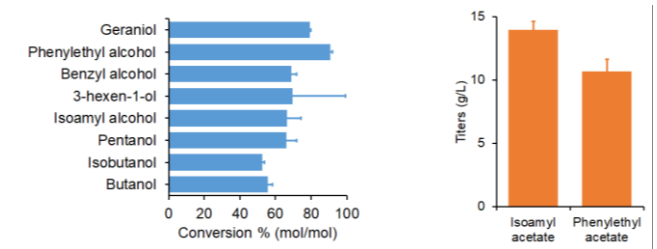
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## Background

- By condensing an acyl-CoA and an alcohol, alcohol acyltransferases (AATs) can serve as an interchangeable metabolic module for microbial biosynthesis of a diverse class of ester molecules with broad applications as flavors, fragrances, solvents, and drop-in biofuels.
- The current lack of robust and efficient AATs limits their utility with precursor pathways and microbial hosts.

## Approach

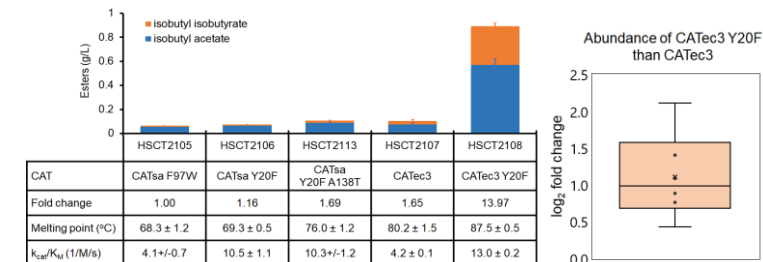
- Through bioprospecting and model-guided protein engineering, we engineered substrate promiscuity of chloramphenicol acetyltransferases (CATs) to function as robust and efficient AATs compatible with at least 21 alcohols and 8 acyl-CoAs for microbial biosynthesis of linear, branched, saturated, unsaturated and/or aromatic esters in mesophiles and thermophiles.



Demonstration of efficiency and compatibility of the engineered AAT in *Escherichia coli* whole-cell biocatalyst

## Outcomes and Impacts

- Developed de novo thermostable AATs that are efficient and compatible with various pathways and microbial hosts (e.g., mesophiles and thermophiles).
- Demonstrated high conversion of various alcohols and achieved about 14 g/L of isoamyl acetate with >95% (mol/mol) conversion efficiency.
- Demonstrated that CAT robustness with enhanced thermostability is critical for efficient ester production in thermophiles by maintaining high level of intracellular CAT abundance.
- Engineered *C. thermocellum* to produce up to 1 g/L of isobutyl esters from cellulose.



Impact of enzyme thermostability in ester production by *C. thermocellum* fermenting cellulose

## Significance

- This work not only presents a robust, efficient, and highly compatible AAT platform for designer bioester production, but also elucidates the impact of enzyme thermostability on engineering heterologous pathways in thermophiles.